THERMAL SYNTHESIS OF THE OPTICAL PURE PENTAPEPTIDE DERIVATIVE Z-(L)-Ala-(L)-Phe-Gly-(L)-Phe-Gly-OMe

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Summary:

The pentapeptide 2-(L)-Ala-(L)-Phe-Gly-(L)-Phe-Gly-OMe was obtained by reacting the activated dipeptide derivative 2-(L)-Ala-(L)-Phe-OPcp with the amino peptide derivative $TFA\cdot H-Gly-(L)-Phe-Gly-OMe$ at 100-105 °C under reduced pressure $(10^{-2}-10^{-3} \text{ Torr})$ without using solvents. The product obtained by bulk condensation showed no racemisation, whereas the product obtained by a matrix mediated condensation contained 5,5 % diastereomer 2-(L)-Ala-(D)-Phe-Gly-(L)-Phe-Gly-OMe. Separation of diastereomers was achieved by HPLC on a silicagel column.

According to Goodman et al.^{1,2}, activated tri- and tetra-peptide esters which are adsorbed on a diatomaeous earth matrix, polymerize without racemization to polypeptides when heated at temperatures of $100-105^{\circ}$ C in high vacuum.

It has been our intention to apply this principle of synthesis to the contruction of defined peptides. One of several model peptides which we have synthesized by this method, and which meets the numerous critèria required for this study has the sequence:

Z-(L)-Ala-(L)-Phe-Gly-(L)-Phe-Gly-OMe.

We chose benzyloxycarbonyl-(L)-alanine-(L)-phenylalanine-pentachlorophenylester (Z-(L)-Ala--(L)-Phe-OPcp) and the trifluoroacetate salt of glycine-(L)-phenylalanine-glycine-methylester (TFA·H-Gly-(L)-Phe-Gly-OMe) as starting compounds for the synthesis.

The reaction followed equation I:

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Equation I: Z-(L)-Ala-(L)-Phe-OPcp + TFA \cdot H-Gly-(L)-Phe-Gly-OMe \rightarrow Z-(L)-Ala-(L)-Phe-Gly-(L)-Phe-Gly-OMe + HOPcp + TFA
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The starting compounds were synthesized by the mixed anhydride method. To ensure that the

dipeptide pentachlorophenyl ester was free of racemization, we employed the "backup route"^{7,2} by coupling HBr \cdot H-(L)-Phe-OPcp with Z-(L)-Ala-OH via the mixed anhydride method. The ester was found to be free of racemization by ¹H-NMR-spectroscopy^{4,5}. The diastereomeric penta-chlorophenyl peptide esters have different chemical shifts for the methyl doublets of alanine (1.0 ppm - 1.1 ppm).

The first attempt at coupling the dipeptide ester with the trifluoroacetate salt of the tripeptide was performed on a diatomacous earth matrix (Hyflo Super Cel^R) at a temperature of 100 - 105°C in a vacuum of $10^{-2} - 10^{-3}$ Torr. After 47 hours only incomplete reaction was obtained. The product C was separated from the matrix by chloroform extraction.

A parallel reaction, a bulk condensation under the same reaction conditions, led within 83 hours to product D. In both cases, the degree of reaction was controlled by TLC. Counter current distribution in a toluene systeme⁸⁾ afforded the products in high purity. The yield was 20 % for the matrix-mediated, and 18 % for the bulk condensation product.

To establish the degree of racemization occuring during the thermal coupling, we synthesized by conventional methods the diastereomeric pentapeptides A and B as references. This permitted us to compare their characteristics with these of the thermal products C and D.

- A) Z-(L)-Ala-(L)-Phe-Gly-(L)-Phe-Gly-OMe
- B) Z-(L)-Ala-(D)-Phe-Gly-(L)-Phe Cly-OMe.

The amount of diastereomer in the pentapeptides C and D, was monitored by optical rotation measurements, and by enzymatic digestion of acid hydrolysate with D-amino acid oxidase^{3,9,10}. The diastereomeric components could also be separated by HPLC, allowing determination of the ratio of the diastereomers by comparison with the standard peptides A and B. (For results see scheme I)

In the evaluation of the enzymatically-determined degree of racemization one must take into account that free <u>L</u>-Phe is partially converted into <u>D</u>-Phe (ca. 2 %) during the acid hydrolysis step⁹. Phenylalanine within peptides may be racemized up to 45 % during the hydrolysis, depending on the neighbouring amino acids⁹! This method, although rapid, can only be used as a rough screening method which allows no useful determinations of low (5 %) degrees of racemization.

The HPLC separation was a baseline separation and was performed on a Perkin Elmer (Series 3) instrument. The column was from Perkin-Elmer, (Silica A, 10 μ m particle size). The eluant was a mixture of chloroform and methanol (100:1,5).

The error in the determination of the <u>D</u>-diastereomer content was reckoned to be 0,3 %, so that lower degrees of racemization than 5 % can be reasonably well determined.

Scheme I:

| Samples: | Analytical data | % Diastereomer, determined by: D-amino acid oxidase | HPLC |
|-----------------------------|---|---|-------------------------------------|
| found: a ²² : | Ala(1):Phe(2): <u>Gly(2)</u> 0,97 1,90 <u>2,00</u> -19,6° <u>+</u> 0,5 c=1 DMF 196 - 197°C | 8 | 0,0 <u>+</u> 0,3 |
| found: α_D^{22} : | Ala(1):Phe(2): <u>Gly(2)</u> 1,08 1,98 <u>2,00</u> -6,1° <u>+</u> 0,5 c=1 DMF 175 - 179°C | 104 | 100,0 <u>+</u> 0,3 |
| α_D^{22} : | Ala(1):Phe(2): <u>Gly(2)</u> 1,00 1,99 <u>2,00</u> -19,1° <u>+</u> 0,5 c=1 DMF 190 - 195°C | -4 | 5,5 <u>+</u> 0,3 |
| found: | Ala(1):Phe(2): <u>Gly(2)</u> 0,99 2,02 <u>2,00</u> -19,9 <u>+</u> 0,5 c=1 DMF | -2 | 0,0 <u>+</u> 0,3 |
| Sample: | Analytical data | method of identification of racemization: | ¹ H-NMR- spectroscopy |
| | Ala(1): <u>Phe(1)</u> 1,00 <u>1,00</u> -67,5°+ 0,5 c=1 DMF | | no racemization |

In summary, it is feasible to use thermal condensation of peptide derivatives for the synthe sis of polypeptides of defined sequence. The use of a matrix, however, leads to increased racemization (11 %) in comparison to that found in bulk condensations where virtually no racemization can be observed.

The low yields of 18 to 20 % could be accounted for side reactions, for example homopolymerisation¹¹ of the amino component or reduced movement of the reacting components. Further investigations are under way to understand the role of side reactions in order to improve the yield of the coupling.

- M. Sakarellos-Diatsiotis, C. Gilon, C. Sakarellos, M. Goodman J. Amer.Chem. Soc. <u>98</u>, 7105 - 7107 (1976)
- 2) D. Nissen, C. Gilon, M. Goodman Makrom. Chem. Suppl. 1, 23 - 53 (1975)
- 3) D.M. Larson, D.C. Snetsinger, P.E. Waibel Anal. Biolog. <u>39</u>, 395 - 401 (1971)
- 4) B. Weinstein, A.E. Pritchard J.C.S. 1015 - 1020 (1972)
- 5) B. Halpern, L.F. Clew, B. Weinstein J. Amer. Chem. Soc. <u>89</u>, 5051 - 5052 (1967)
- 6) M. Goodman, L. Levin J. Amer. Chem. Soc. <u>86</u>, 2918 - 2922 (1964)
- 7) M. Goodman, K.C. Stueben J. Amer. Chem. Soc. <u>81</u>, 3980 - 3983 (1959)
- 8) C.M. Li, J. Meienhoffer, E. Schnabel, J.B. Lo, J. Ramachandran J. Amer. Chem. Soc. 82, 5760 - 5762 (1960)
- 9) J. Gebhardt, H. Zahn Diploma Thesis, Technical University Aachen, (1979)
- 10) E. Jäger Max-Planck-Institut für Biochemie, München private communication
- 11) G. Schramm, H. Restle Makrom. Chem. 13, 103 -116 (1954)

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